

A genetic model for gallbladder carcinogenesis and its dissemination

S. G. Barreto¹, A. Dutt^{2*} & A. Chaudhary¹

¹Department of Gastrointestinal Surgery, Gastrointestinal Oncology, and Bariatric Surgery, Medanta Institute of Digestive and Hepatobiliary Sciences, Medanta, The Medicity, Gurgaon; ²The Advanced Centre for Treatment, Research and Education in Cancer, Tata Memorial Centre, Kharghar, Navi Mumbai, India

Received 31 August 2013; revised 5 November 2013; accepted 8 November 2013

Gallbladder cancer, although regarded as the most common malignancy of the biliary tract, continues to be associated with a dismal overall survival even in the present day. While complete surgical removal of the tumour offers a good chance of cure, only a fraction of the patients are amenable to curative surgery owing to their delayed presentation. Moreover, the current contribution of adjuvant therapies towards prolonging survival is marginal, at best. Thus, understanding the biology of the disease will not only enable a better appreciation of the pathways of progression but also facilitate the development of an accurate genetic model for gallbladder carcinogenesis and dissemination. This review provides an updated, evidence-based model of the pathways of carcinogenesis in gallbladder cancer and its dissemination. The model proposed could serve as the scaffolding for elucidation of the molecular mechanisms involved in gallbladder carcinogenesis. A better understanding of the pathways involved in gallbladder tumorigenesis will serve to identify patients at risk for the cancer (and who thus could be offered prophylactic cholecystectomy) as well as aid oncologists in planning the most suitable treatment for a particular patient, thereby setting us on the vanguard of transforming the current treatment paradigm for gallbladder cancer.

Key words: gallbladder cancer, genetic alteration, carcinogenesis, dissemination, biology, therapeutics

introduction

Gallbladder cancer is relatively uncommon worldwide with age-standardized incidence rates of 2/100 000 [1]. Besides, recent reports have suggested a decline in its incidence in different parts of the world [2, 3]. However, in Chile and India, gallbladder cancer remains a major problem [4, 5] (age-standardized rates from 3.9 to 8.6/100 000 [6]) with the vast majority of patients presenting with advanced disease [7, 8]. The uncommonness of gallbladder cancer has contributed to the generally poor understanding of the disease [9, 10]. However, despite the suggested declining trend of this cancer in the world, it is imperative that a better understanding of the disease and the factors influencing its course is needed to develop treatment strategies aimed at improving its overall outcome.

The most important strategy to successfully plan treatment options for gallbladder cancer is to first understand the pathogenesis of the disease. A useful tool to this end is the development of a comprehensive model of carcinogenesis akin to the Fearon–Vogelstein model for colorectal tumorigenesis [11]. By developing such a model (that will incorporate stages before the formation of an invasive cancer and up until tumour dissemination), it will be possible to lay the ground-work for a more

dedicated thrust towards evidence-based, targeted initiatives in the management of gallbladder cancer. This review provides a road-map towards the development of such a model.

can we equate gallbladder cancer with colorectal cancer in terms of a natural evolution of carcinogenesis?

To even consider embarking on a tumorigenesis model, it is important to first ascertain if gallbladder cancer development progresses from benign to malignant in a step-wise evolution similar to colorectal cancer (adenoma to carcinoma).

The work by Laitio [12, 13] provided a basis for the understanding of the stages in the pathogenesis of gallbladder cancer that would eventually lead to the elucidation of a step-wise progression. Laitio [13] demonstrated that metaplasia in the gallbladder wall could develop into dysplasia which could play a significant role in gallbladder carcinogenesis [12]. The earliest work hinting at the existence of a step-wise evolution of gallbladder cancer by Albores-Saavedra et al. [14] suggested that hyperplasia could potentially develop into atypical hyperplasia and from there on to *in situ* and finally invasive cancer. The quest for a natural evolution in gallbladder carcinogenesis was further placed on a firm footing by the work of Roa et al. [15] who not only suggested the dysplasia–carcinoma sequence but also indicated that the time to transformation from dysplasia to

*Correspondence to: Dr Amit Dutt, Wellcome Trust/DBT India Alliance Intermediate Fellow, Tata Memorial Centre, ACTREC, Navi Mumbai 410 210, India. Tel: +91-22-27405056; E-mail: adutt@actrec.gov.in

advanced carcinoma was ~15 years. Roa et al. [16] went on to eloquently demonstrate the presence of metaplasia, dysplasia and *in situ* carcinoma in the vicinity of the invasive carcinoma lending further support to the metaplasia–dysplasia–carcinoma cascade first postulated by Yamagiwa and Tomiyama [17].

Duarte et al. [18] on extensive histological analysis of gallbladder benign and malignant specimens noted a significant association between intestinal metaplasia, hyperplasia and dysplasia.

The above work, however, largely originated from South and Central America [19]. In these countries, as is the case in India and most parts of the world, cholelithiasis is regarded as a cause, if not a co-factor, in the development of cancer [5]. Moreover, the p53 mutation occurs as an early event [20, 21] in these regions and is associated with gallstone disease.

An appreciation of yet another distinct pathway of disease progression was put forth by Kozuka et al. [22] who noted the presence of large adenomas (>12 mm) in relation to gallbladder cancer. This was later followed by the work of Watanabe et al. [23] suggesting three distinct pathways in gallbladder carcinogenesis—depending on the predominance of K-ras or p53 mutations.

According to Nakajo et al. [24], adenomas of either the metaplastic type or non-metaplastic type could progress to adenocarcinomas.

It thus emerged that there existed two pathways in the pathogenesis of gallbladder cancer, viz: the dysplasia–carcinoma sequence in patients with gallstones; and the adenoma–carcinoma cascade [25]. However, of the two, the more plausible cascade remains the dysplasia–carcinoma sequence [26] owing to the generally low incidence of adenomas of the gallbladder and their co-existence in the vicinity of early or advanced cancer [26] as well as some evidence to suggest different pathways

being involved in adenoma development when compared with adenocarcinoma [27].

In Japan and the Far East, a morphological biliary anomaly termed anomalous pancreaticobiliary ductal junction (APBDJ) is associated with gallbladder development in a much higher frequency when compared with the rest of the world [28–30]. In APBDJ, hyperplasia is noted in up to 61% [25] of individuals. It has been postulated that hyperplasia is more likely to develop into an invasive cancer through the dysplasia–carcinoma cascade.

Thus, it is certainly possible to develop a multi-step evolutionary sequence from normal mucosa to malignant change in gallbladder cancer just as was done by Fearon and Vogelstein [11] for colorectal cancer. Only the dominant dysplasia–carcinoma tumorigenesis sequence will be discussed further.

the multi-step pathogenesis of gallbladder carcinogenesis (dysplasia to carcinoma)

In 1999, Wistuba and Albores-Saavedra [31] provided the first image of the dysplasia–carcinoma cascade based on sequential histopathological and molecular changes in the pathogenesis of gallbladder carcinoma associated with gallstones and inflammation. In 2004, Wistuba and Gazdar [32] improved on this by providing the median ages at diagnosis of each histopathological change. Figure 1 provides a diagrammatic representation of the Wistuba and Gazdar multi-step pathogenetic sequence.

While this marked the first representation of the cascade, one of the lacunae was the omission of stage of metaplasia. The importance of the metaplastic change cannot be undermined in gallbladder cancer, especially considering that >80% are

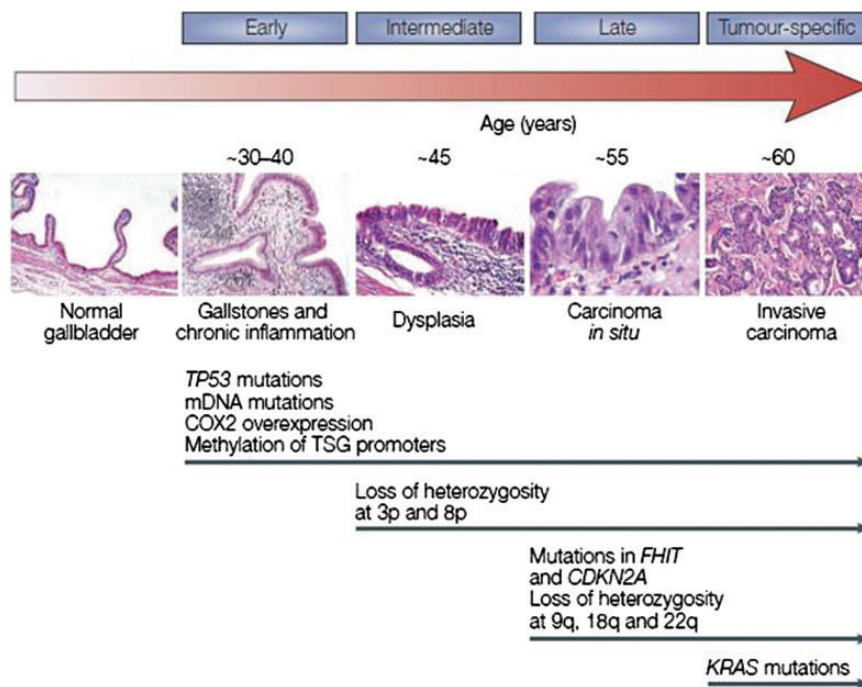


Figure 1. Multi-step pathogenesis of gallbladder cancer from gallstones proposed by Wistuba and Gazdar’s [32]. TSG, Tumour suppressor gene; FHIT, fragile histidine triad; mDNA, mitochondrial deoxyribonucleic acid; COX, cyclooxygenase.

adenocarcinomas. This is ironic when one considers that the overriding putative inciting factor is chronic irritation by gallstones [5]. Mere irritation should have resulted in a higher proportion of squamous differentiations rather than adenocarcinoma. More recently, Castillo et al. [33] provided an update of the genetic alterations in gallbladder cancer on the template of the 'metaplasia–dysplasia–carcinoma' sequence first postulated by Yamagiwa and Tomiyama [17].

where do we go from here?

The two cascades in the development of gallbladder cancer proposed have been based on laboratory evidence of some markers of genetic and epigenetic alterations. However, although they form the basis for any further development of the tumorigenesis model, they remain a work in progress.

Two prime alterations that need to be incorporated into the existing models are:

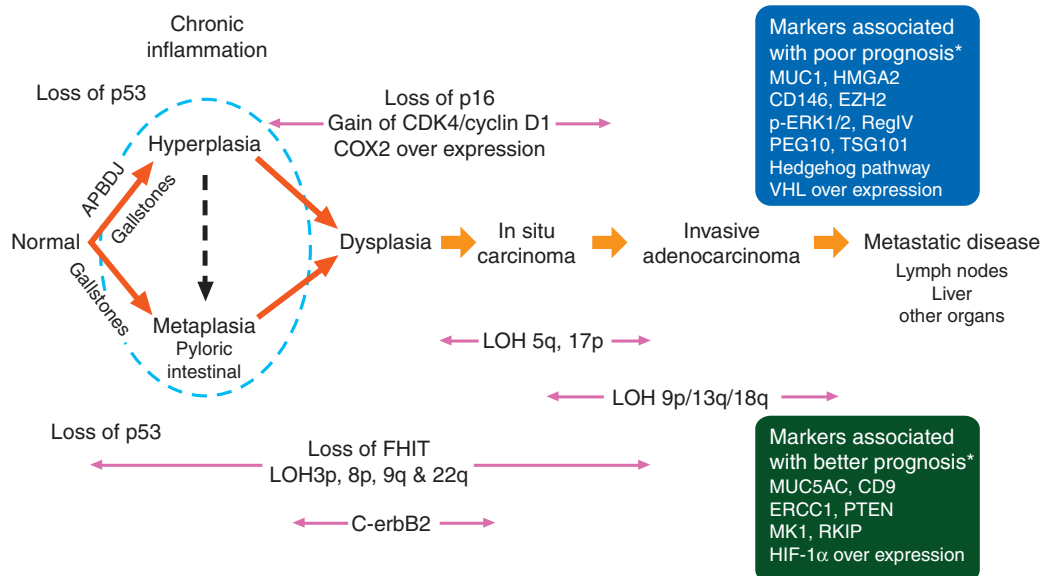
- Course of the disease after the development of invasive carcinoma within the gallbladder, namely tumour dissemination to lymph nodes, liver and other organs.
- Updating the existing chart with markers that may be specific to the precancerous stage, invasive stage or stage of dissemination—markers which could be used in screening, diagnosis, guide treatment—choice of therapy or response to therapy.

This will permit the development of a 'working' tumorigenesis model that will enable us to better understand the disease in its entirety, thereby serving as a blue-print for planning screening/surveillance and management strategies.

Based on these principles, an updated model is presented below—'The Gallbladder Carcinogenesis and Dissemination Model' based on a review of literature with an aim to update the pre-existing cascades [25, 32, 33].

the 'gallbladder carcinogenesis and dissemination model'

The skeletal framework of this model is based on evidence indicating that primal to the development of gallbladder cancer is the existence of chronic inflammation (Figure 2) [34]. Chronic inflammation results either due to gallstones or due to changes in the bile owing to the reflux of pancreatic juice into the common bile duct induced by APBDJ [35, 36]. Chronic inflammation secondary to gallstones is more likely to lead to metaplasia (intestinal or pseudopyloric) [18] rather than hyperplasia [14], while the cholecystitis due to APBDJ on the other hand is more likely to lead to hyperplasia [37]. Further, dysplasia has been noted to develop in metaplastic epithelium [13] as well as from hyperplastic epithelium that has progressed to the atypical type [14]. It is likely that hyperplasia (that has developed at an early onset in patients with APBDJ) could also progress to



* For the exception of MUC1 & MUC5AC, the other markers are based on single studies and merit further validation

Figure 2. The 'Gallbladder carcinogenesis and dissemination model'. The proposed model takes into consideration each and every pathological change occurring in the gallbladder epithelium progressing sequentially from normal epithelial mucosa to the development of cancer via the two most common pathways, namely metaplasia/hyperplasia as well as dysplasia, and beyond the localized disease in the gallbladder to even include the spread of the cancer to regional and distant organs. The putative molecular alterations playing a role in each step are also highlighted. APBDJ, anomalous pancreaticobiliary ductal junction; FHIT, fragile histidine trait; LOH, loss of heterozygosity; HMGA2, high mobility group protein A2; CD9, mobility related protein 1; CD146, melanoma cell adhesion molecule; ERCC-1, excision repair cross-complementing group 1; EZH2, histone-lysine *N*-methyltransferase; PTEN, phosphatase and tensin homologue; pERK1/2, extracellular signal-regulated kinase; PI3-K, phosphatidylinositol-3 kinase; RKIP, Raf-1 kinase inhibitory protein; RegIV, member of regenerating gene family; MK-1, type 1 transmembrane protein (Ep-CAM); PEG10, retrotransposon-derived protein 10; TSG101, tumour susceptibility gene 101; VHL, Von Hippel–Lindau gene; HIF1 α , hypoxia-inducible factor 1 α ; CD, cluster of differentiation; COX, cyclooxygenase.

dysplasia via the stage of metaplasia [25]. However, this is simply an assumption based on the significant co-occurrence of metaplasia and hyperplasia [18] that remains as yet unconfirmed. The postulate that dysplasia is a premalignant lesion that potentially develops into carcinoma *in situ* and invasive adenocarcinoma is derived from two important aspects [16], namely the consistent finding of dysplasia in the vicinity of gallbladder cancer more frequently than in non-malignant tissue on histological examination (88% versus 34%) [25, 38] and the appreciation of similar genetic alterations in dysplastic and malignant gallbladder tissue [31, 39]. After the development of invasive carcinoma of the gallbladder, the two most common sites for metastases are the lymph nodes and the liver [40].

Important to note in the above model is the non-inclusion of the adenoma–carcinoma cascade. This does not imply that the authors do not accept the existence of this cascade but rather opted to focus on the more common pathway involved in gallbladder carcinogenesis based on evidence from world literature [41].

genetic alterations in the development and dissemination of gallbladder cancer

p53

The tumour suppressor gene p53 has been found to occur in patients with gallbladder cancer throughout the world [20, 21, 42], although differences in the mutational spectra have been

reported in tumour specimens from Japan and Chile (Figure 3) [43]. The most common acquired mutations are in exons 5 and 8. p53 mutations constitute one of the earliest changes in the development of gallbladder cancer—being detected in one-third of normal and dysplastic epithelia obtained from gallbladders with gallstones but without cancer [44]. Wistuba et al. [45] noted that the loss of heterozygosity (LOH) of p53 occurred earlier, and more frequently, than protein over-expression.

p16/cyclin d1/CDK4

In a recent study, Feng et al. [46] addressed the role of the p16/cyclin D1/CDK4 pathway in gallbladder cancer via the hyperplasia pathway. They noted that while expressions of CDK4 and cyclin D1 increased along with the progression of gallbladder mucosa hyperplasia with the highest expression noted in the cancer group, p16 decreased to the lowest level in gallbladder cancer. This loss of expression has been summarized by Goldin and Roa [26] to be either due to deletions in region 9p21, inactivation of the gene, LOH or by methylation.

KRAS

The evidence available is confounding with studies suggesting little or no role of the oncogene KRAS in gallbladder cancers [27, 45, 47, 48], one study reporting the detection of KRAS mutations in 59% of cancers and 73% of gallstone-induced dysplasia, while others indicating the presence of the mutation in a proportion of cancers arising secondary to APBDJ [49, 50]. The study by Itoi

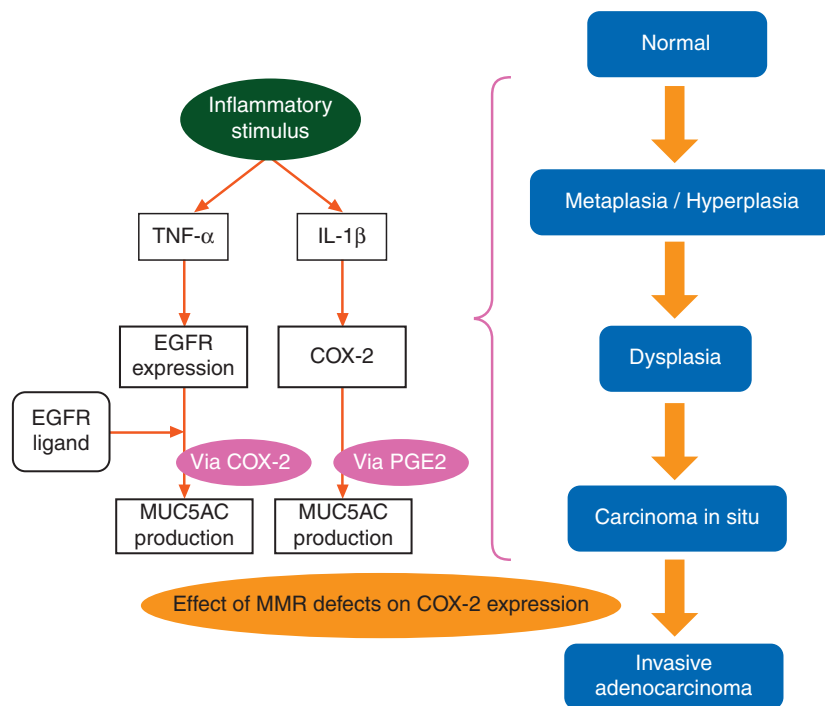


Figure 3. Inflammatory cascade hypothesis in relation to the development of gallbladder cancer. This is as yet a hypothetical model for gallbladder carcinogenesis via inflammation and inflammatory markers derived from observations in studies examining the expression of these markers in the various stages of gallbladder carcinogenesis. The model is built deriving inferences from observations in colon cancer and gastric and airway epithelial cells and cell cultures analysing the impact of inflammatory markers. TNF- α , tumour necrosis factor- α ; EGFR, epidermal growth factor receptor; COX-2, cyclooxygenase-2; PGE-2, prostaglandin E2; IL-1 β , interleukin-1 β ; MMR, mismatch repair [95, 112–114].

et al. [51] analysing the status of KRAS mutations in gallbladder cancer suggests that the dominant pathway for gallbladder cancer pathogenesis may not involve KRAS mutations.

cyclooxygenase-2

Cyclooxygenase-2 (COX-2) over-expression occurs early in the pathogenetic cascade of gallbladder carcinogenesis being detected in high proportions in dysplasia and invasive carcinoma when compared with normal epithelium [52]. COX-2 expression has actually been noted to be reduced in adenocarcinoma tissue (59.2%) when compared with dysplastic epithelium (70.3%). COX-2 expression has also been noted to be significantly higher in epithelial hyperplasia secondary to APBDJ when compared with normal epithelium [53]. In malignant tissue, the expression of COX-2 is significantly lower in the histopathologically normal surrounding epithelium [54].

deleted in colorectal carcinoma: 18q21

LOH has been noted at deleted in colorectal carcinoma (DCC) in gallbladder cancers [45, 55] and recognized to be an early event in carcinogenesis [45]. Genome-wide association studies have recently demonstrated that a specific DCC haplotype is associated with increased susceptibility to gallbladder cancer in India, irrespective of associated risk factors [56]. A similar study reported single-nucleotide polymorphisms in DCC associated with gallbladder cancer in a Japanese population [57].

fragile histidine triad gene

Wistuba et al. [58] studied the frequency of loss and LOH at fragile histidine triad (FHIT) in normal, dysplastic and malignant tissue. They noted that while occasional FHIT abnormalities were occasionally demonstrated in histologically normal epithelium, the reduction or absence of FHIT immunostaining significantly reduced with evidence of disease progression through dysplasia and to adenocarcinoma. They noted a high correlation between immunostaining in the specimen and allelic loss. This finding of a reduction in expression of FHIT in gallbladder cancer was also noted by Koda et al. [59].

LOH of other chromosomes

As per Knudson's hypothesis [60], LOH at polymorphic loci is recognized as a hallmark of a tumour suppressor gene whose other allele is inactivated by point mutations or by some other mechanism [61]. Chang et al. [62] studied LOH on chromosomal regions 3p, 5q, 8p, 9p, 13q, 17p and 18q in gallbladder specimens with dysplasia and carcinoma. They noted that while LOH on 5q was an early change of carcinogenesis of the gallbladder, LOH on 3p and 9p was related to the progression of gallbladder carcinoma with LOH on 13q and 18q likely to be late events. They also noted that LOH on 17p occurred not only in dysplasia but also increased during the subsequent stages. On the other hand, Wistuba et al. [61] noted that LOH was an early phenomenon in the development of cancer since they found increasing proportions of LOH on chromosomes 3p, 8p, 9q and 22q in normal epithelium, dysplastic and malignant tissue.

microsatellite instability

Mismatch repair (MMR) gene mutations [63, 64] and the resultant microsatellite instability (MSI) are infrequently detected in gallbladder cancer [55, 62, 65]. An interesting perspective on this has been presented by Matsuda [66] who suggested that the association of MSI-positive tumours harbouring a favourable prognosis may imply a similarity to colon cancer. He further pointed out that the difference in the two cancers in terms of carcinogenesis may lie in the fact that gallbladder cancers tended to be MSI-L, while colon cancers were more likely to be MSI-H.

c-erb b2/HER2

The HER2 protein expression was recently studied in normal, metaplastic and invasive adenocarcinoma tissue, as well as in samples of carcinoma *in situ* [67]. Interestingly, while normal epithelium failed to show any HER2 immunoreactivity, maximal immunoreactivity was noted in metaplastic tissue (intestinal) and in tissue from carcinoma *in situ*. Once again the immunoreactivity dropped in invasive cancer. The findings of Toledo et al. [67] are interesting as before this, Kamel et al. [68] and Kim et al. [69] had failed to demonstrate HER2 expression in dysplastic tissue while demonstrating a similar expression in malignant tissue. Chaube et al. [70] found that the expression levels for HER2 varied depending on the grade of the tumour—with decreasing expression correlating with advancing grade.

markers studied in normal, premalignant and malignant tissues of gallbladder adenocarcinoma

epidermal growth factor receptor

Studies examining the expression of epidermal growth factor receptor (EGFR) in gallbladder cancer have indicated a highly variable expression of the receptor (Table 1 [71–89]). The expression ranged from 11.3% to 100% [75, 77, 78, 80–82, 85]. The major impediment to the understanding of the role of EGFR in gallbladder carcinogenesis has been the lack of studies examining the expression profile simultaneously in the premalignant tissue samples. Kim et al. [90] noted a reduced EGFR expression in gallstone patients when compared with normal controls. EGFR and Her2 belong to a family of receptor tyrosine kinases that are anchored in the cytoplasmic membrane and share a similar structure [91]. The expression profile of Her2, as noted above, is not uniform throughout the process of gallbladder carcinogenesis. Thus, future studies in which a comparative expression profile of the expression of EGFR from normal gallbladder tissue to metaplasia/hyperplasia to dysplasia and further to *in situ* and invasive adenocarcinoma would aid in our understanding of the role of EGFR in gallbladder cancer.

MUC

Mucins are major components of the mucous viscous gel lining epithelial tissue surfaces [92]. Xiong et al. [87] reported that MUC staining on immunohistochemistry was noted mainly in the cytoplasm and/or the cell membrane rather than the nucleus. Of the nine distinct epithelial mucin genes identified in gallbladder diseases, MUC1, MUC5AC and MUC6 are normally expressed in

Table 1. Studies investigating novel alterations in normal, premalignant and malignant gallbladder cancer [71–88]

Marker	Normal	Hyperplasia	Metaplasia	Dysplasia	Carcinoma <i>in situ</i>	Invasive cancer	References
EGFR	—	—	—	—	—	38.4% (strong)	[75]
	—	—	—	—	—	83% (overall)	[85]
	—	—	—	—	—	33% (strong)	
	—	—	—	100% (moderate)	—	100% (strong)	[78]
	—	—	—	—	—	12.4% (strong)	[81]
	—	—	—	—	—	93.7% (overall)	[77]
	—	—	—	—	—	75% (strong)	
	—	—	—	—	16.6% (strong)	11.4% (strong)	[80]
	—	—	—	—	—	11.3% (strong)	[82]
	CK 7	100%	—	100%	100%	—	87%
100%		100%	100%	—	—	100%	[71] ^a
—		—	—	—	—	69.50%	[76]
—		—	—	—	—	82%	[74]
CK20	0%	—	17%	31%	—	18%	[72]
	0%	100%	100%	—	—	100%	[71] ^a
	—	—	—	—	—	28.50%	[76]
	—	—	—	—	—	27%	[74]
MUC1	0%	—	0%	35%	—	75%	[72]
	—	—	—	21.70%	—	57.40%	[87]
	—	—	—	50%	—	80%	[88]
	0–20%	—	—	—	—	78–89%	[89]
MUC2	0%	—	29%	9%	—	11%	[72]
	—	—	—	75%	100%	58%	[84]
	—	—	—	75%	—	64%	[88]
	—	—	91.70%	—	—	—	[83]
MUC5AC	89%	—	92%	53%	—	38%	[72]
	21.70%	—	—	60.80%	—	51.90%	[87]
	—	—	—	85%	90%	78%	[84]
MUC6	100%	—	100%	65%	—	27%	[72]
	—	—	—	80%	90%	91%	[84]
CDX2	—	—	—	—	—	29.20%	[73]
	—	—	91.70%	—	—	—	[83]
	0%	—	—	—	—	36.80%	[86]
	—	100%	—	—	—	45.40%	[79]

^aIndividual break-down of the cases was not provided.

EGFR, epidermal growth factor receptor; CK, cytokeratin; MUC, mucin; Strong => ≥2+.

gastric mucosa, while MUC2 is expressed in intestinal mucosa [93]. Thus, an extrapolation of this mucin expression profile in gastric cancer [93] would lead to the assumption that in gallbladder cancer developing from pyloric metaplasia, MUC1, MUC5AC and MUC6 would exhibit a rather similar expression profile, while MUC2 over-expression would predominate in cancers developing from intestinal metaplasia. However, contrary to this, the expression of MUC in gallbladder cancers is not uniform. MUC1 levels are significantly increased in gallbladder cancer, MUC2 and MUC5AC expression levels are reduced in gallbladder cancer when compared with dysplastic tissue as well carcinoma *in situ*, while the findings in the case of the two studies reporting expression levels of MUC6 are contradictory [72, 83, 84, 87–89]. The localization of MUC1 on chromosome locus 1q21 and MUC2, MUC5AC and MUC6 on chromosome locus 11p15.5 [94] may be a potential reason for this differential expression.

Finzi et al. [95] have suggested that the MUC5AC is overproduced in gallstone disease by an inflammation-dependent EGFR cascade. Vilkin et al. [96] noted that the extent of MUC5AC expression as a result of inflammation was more in pigment gallstones.

Xiong et al. [87] found that the expression of MUC1 and MUC5AC were inversely related when correlated with the extent of disease. Lower MUC1 and higher MUC 5AC expression levels were noted in tumours that were <2 cm, with no lymph node or regional tissue involvement. Also, the expression levels of MUC1 were significantly higher in tumour when compared with peritumoural tissue, while the converse was true in the case of MUC5AC. Chang et al. [72], too, appreciated the correlation of MUC1 with more aggressive tumours, while Ghosh et al. [72, 83, 84, 87–89] noted that its depolarized expression was a marker of invasion.

The above findings of the expression levels of MUC5AC in gallbladder cancer are exactly the opposite as reported in cholangiocarcinoma. Boonla et al. [97] and Park et al. [98] noted that in cholangiocarcinoma, serum MUC5AC levels were predictive of poor outcomes.

cytokeratins

Cytokeratins have been explored as markers for detecting micrometastatic disease in lymph nodes previously reported as normal on histopathology [99–101]. Micrometastatic disease is an indicator of poorer outcomes [100].

other markers

Recently, there have been numerous publications investigating markers, previously shown to be involved in other solid organ cancers, in gallbladder adenocarcinomas. Table 2 provides an overview of these studies [102–111]. These markers need validation in larger series to confirm their usefulness as prognostic markers.

the inflammatory cascade and marker expression

An interesting finding in gallbladder cancer, as noted above, is the elevation of inflammatory cascade markers such as EGFR, MUC5AC and COX-2, early in gallbladder carcinogenesis up until the stages of dysplasia and even *in situ* carcinoma followed by a reduction in their expression in invasive adenocarcinoma. This observation raises an important question—if inflammation

does play a role in the development of gallbladder cancer from gallstones, then why do the levels of these markers, which are important in inflammation, undergo a reduction between the stages of dysplasia to invasive adenocarcinoma? Could these markers be actually exerting a protective influence (via the production of protective mucin through MUC5AC), while the overcoming/overwhelming of these protective forces results in invasive cancer?

Figure 3, based on the work in colon cancer and gastric epithelial cells [112] and human airway epithelial cells and cell cultures [113, 114], lends support to the first portion of the hypothesis, namely inflammation and inflammatory markers in gallbladder cancer (tumour necrosis factor- α /TNF- α and interleukin-1 β /IL-1 β [115]) leading to the constitutive expression of protective mucin—MUC5AC [95] via an EGFR and COX-2-mediated pathway. Patients with hereditary non-polyposis colorectal cancer whose basic genetic defect lies in the MMR genes appear to have a reduced expression of COX-2 in their tumours when compared with those individuals with sporadic colorectal cancers [116, 117]. Defects in MMR genes have been noted to increase an individual's susceptibility to gallbladder cancer [63]. Moreover, loss of O⁶-methylguanine-DNA methyltransferase, a DNA repair enzyme, as well as MMR proteins (hMLH1 and hMSH2) were associated with a poor prognosis in gallbladder cancer [64]. MSI has been noted in gallbladder cancer. Thus, one potential explanation for the loss of protective mucin MUC5AC in the later stages of carcinogenesis could be the result of defects in MMR leading to a reduced expression of COX-2 and hence MUC5AC.

how can we further improve the model to strengthen its clinical relevance?

Variations in mutation frequency in gallbladder cancer have been attributed to geographical and ethnic variability in the disease. Non-uniformity of technique between different laboratories could also be contributory [118]. However, the appreciation of tumour clonality and tumour heterogeneity in solid organ cancers [119, 120] adds a new dimension to the perceived 'variations in reported mutations'. We have no reason to not believe that gallbladder cancer, too, would exhibit clonality and tumour heterogeneity. The advances in cancer genomics (next-generation sequencing) will most likely aid in delivering critical insights into all stages of tumour progression establishing additional genetic determinants driving the process of carcinogenesis. Cutting edge technology and informatics can potentially elucidate the steps in carcinogenesis quicker, more efficiently and possibly even cost-effectively, making it possible to characterize the biology of primary as well as recurrent gallbladder cancer to an extent far surpassing the extent of our current knowledge. Using contemporary technologies, one can go both deep and wide into cancer genomics with whole-genome studies, targeted gene profiling, gene expression and epigenetic analysis.

implications of developing a successful model

patients with an inherited risk of gallbladder cancer

Gallbladder cancer is known to occur as part of hereditary cancer syndromes such as Lynch syndrome [121], neurofibromatosis 1

Table 2. Individual studies reporting novel alterations in premalignant and malignant tissues of gallbladder cancer along with their proposed significance [102–111]

Marker	Normal	Chronic cholecystitis	Adenocarcinoma	Feature	Reference
HMGA2	—	14.3%	59.3%	Directly correlated with size of the tumour (>2 cm), lymph node metastases, poorer differentiation and regional tissue invasion	[110]
CD9	—	88.6%	52.8%	Inverse correlation with size of the tumour (>2 cm), lymph node metastases, poorer differentiation and regional tissue invasion	[110]
CD146	—	5.7%	53.7%	Directly correlated with size of the tumour (>2 cm), lymph node metastases, poorer differentiation and regional tissue invasion	[107]
ERCC1	95%	—	53%	ERCC1 expression correlated with better differentiation and in subserosal (T2) tumours, ERCC1 staining was associated with a better survival	[111]
EZH2	—	0%	53.7%	EZH2 over-expression is associated with poor prognosis	[105]
PTEN	—	100%	48.2%	PTEN loss of expression is associated with poor prognosis	[105]
pERK1/2	—	11.4%	58.3%	p-ERK1/2 over-expression correlated with decreased survival	[104]
PI3-K	—	8.6%	50.9%	PI3K may contribute to gallbladder carcinogenesis	[104]
Hedgehog pathway				Hedgehog pathway is frequently expressed in gallbladder cancer and is associated with poorer survival	[103]
Shh	0%	—	81.7%		
Ptch1	0%	—	75.3%		
Gli1 protein	0%	—	70%		
RKIP	—	100%	57.7% (tumour); 31.2% (lymph node)	Loss of RKIP may contribute to tumour invasiveness and metastasis and is associated with reduced survival	[102]
RegIV	—	11.4%	53.7%	Directly correlated with lymph node metastases, poorer differentiation, and regional tissue invasion	[108]
MK1	—	14.3%	62%	Inverse correlation with lymph node metastases, poorer differentiation, and regional tissue invasion	[108]
PEG10	—	5.7%	48.1%	Directly correlated with lymph node metastases, poorer differentiation, regional tissue invasion and poorer survival	[106]
TSG101	—	5.7%	47.2%	Directly correlated with lymph node metastases, poorer differentiation, regional tissue invasion and poorer survival	[106]
VHL	—	88.6%	48.1%	Positive expression of VHL significantly associated with differentiation, tumour mass, lymph node metastasis and invasion of adenocarcinoma	[109]
HIF1 α	—	14.3%	53.7%	Negative expression of HIF-1 α significantly associated with differentiation, tumour mass, lymph node metastasis and invasion of adenocarcinoma	[109]

HMGA2, high mobility group protein A2; CD9, mobility-related protein 1; CD146, melanoma cell adhesion molecule; ERCC-1, excision repair cross-complementing group 1; EZH2, histone-lysine *N*-methyltransferase; PTEN, phosphatase and tensin homologue; pERK1/2, extracellular signal-regulated kinase; PI3-K, phosphatidylinositol-3 kinase; Shh, sonic hedgehog; Ptch1, Shh receptor–Patched; Gli1 protein, Shh downstream transcription factor; RKIP, Raf-1 kinase inhibitory protein; RegIV, member of regenerating gene family; MK-1, type 1 transmembrane protein (Ep-CAM); PEG10, retrotransposon-derived protein 10; TSG101, tumour susceptibility gene 101; VHL, Von Hippel–Lindau gene; HIF1 α , hypoxia-inducible factor 1 α .

[122] and Gardner's syndrome [123]. A nation-wide study from Sweden [124] concluded that the risks of familial clustering of gallbladder cancer cases were so high to suggest a contributory role, albeit modified by environmental factors. They noted that demonstration of candidate genes would help further characterize the familial risks. Developing a complete understanding of the genetic transformations involved in gallbladder carcinogenesis could aid in selecting markers for screening the disease, especially in regions of high incidence of gallbladder cancer. It can only be conjectured at this point that the determination of such markers could guide decision-making regarding prophylactic cholecystectomy in mutation carriers.

patients with surgically resectable gallbladder cancer

The development of prognostic markers could aid in deciding which patients with node-negative disease would benefit from adjuvant treatment. Besides, decisions on the choice of therapy in patients with node-positive disease could be decided based on biomarkers.

patients with metastatic gallbladder cancer

Identification of prognostic markers could aid in deciding which patients would benefit from specific palliative strategies (chemotherapy, radiotherapy and/or targeted therapies), thereby permitting the optimal use of resources and finances. Traditionally, the management of advanced gallbladder cancer involved chemotherapeutic agents like gemcitabine and 5-fluorouracil. However, recent reports on the potential role of targeted therapies [125–127] using anti-angiogenic, anti-HER-2/neu or novel MAPK/ERK kinase (MEK) inhibitors lends further support to the concept of the tumorigenesis model to permit a better approach to developing treatment strategies.

This review provides an up-to-date, evidence-based model of gallbladder carcinogenesis and its dissemination. It serves as the scaffolding for the eventual complete elucidation of the exact mechanisms involved in gallbladder carcinogenesis. We foresee the development of molecularly based individualized cancer care as a result of further elucidation of this tumorigenesis pathway using conventional as well as broad-based genomic platforms. Such an approach could not only enable, but reinforce, a cyclical process of selecting treatment for an individual patient based on the genetic expression, proteomic profiles, deregulated cellular pathways and/or somatic mutations in cancer cells of each individual patient, using this profile to accurately define the prognosis in these patients, and suggesting treatment options or clinical trials that are most likely to succeed—something that can be related with the pathological heterogeneity in clinical response frequently observed in clinics, thereby setting us on the vanguard of transforming the current cancer treatment paradigm.

funding

AD is supported by an Intermediate Fellowship from the Wellcome Trust/DBT India Alliance (IA/I/11/2500278), by a grant from DBT (BT/PR2372/AGR/36/696/2011) and intramural grants (Seed-In-Air 2897, TMH Plan Project 2712 and IRB 92).

disclosure

The authors have declared no conflicts of interest.

references

1. Ferlay J, Shin HR, Bray F et al. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 2010; 127: 2893–2917.
2. Alexander S, Lemmens VE, Houterman S et al. Gallbladder cancer, a vanishing disease? *Cancer Causes Control* 2012; 23: 1705–1709.
3. Le MD, Henson D, Young H et al. Is gallbladder cancer decreasing in view of increasing laparoscopic cholecystectomy? *Ann Hepatol* 2011; 10: 306–314.
4. Dhir V, Mohandas KM. Epidemiology of digestive tract cancers in India IV. Gall bladder and pancreas. *Indian J Gastroenterol* 1999; 18: 24–28.
5. Shrikhande S, Barreto S, Singh S et al. Cholelithiasis in gallbladder cancer: coincidence, cofactor, or cause! *Eur J Surg Oncol* 2010; 36: 514–519.
6. Curado M, Edwards B, Shin H et al. Cancer Incidence in Five Continents. IARC Scientific Publications, 2007; IX.
7. Shukla PJ, Neve R, Barreto SG et al. A new scoring system for gallbladder cancer (aiding treatment algorithm): an analysis of 335 patients. *Ann Surg Oncol* 2008; 15: 3132–3137.
8. Doval D, Sekhon J, Gupta S et al. A phase II study of gemcitabine and cisplatin in chemotherapy-naïve, unresectable gallbladder cancer. *Br J Cancer* 2004; 90: 1516–1520.
9. Boutros C, Gary M, Baldwin K et al. Gallbladder cancer: past, present and an uncertain future. *Surg Oncol* 2012; 21: e183–191.
10. Shrikhande SV, Barreto SG. Surgery for gallbladder cancer: the need to generate greater evidence. *World J Gastrointest Surg* 2009; 1: 26–29.
11. Fearon E, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; 61: 759–767.
12. Laitio M. Early carcinoma of the gallbladder. *Beitr Pathol* 1976; 158: 159–172.
13. Laitio M. Histogenesis of epithelial neoplasms of human gallbladder I. Dysplasia. *Pathol Res Pract* 1983; 178: 51–56.
14. Albores-Saavedra J, Alcántra-Vazquez A, Cruz-Ortiz H et al. The precursor lesions of invasive gallbladder carcinoma. Hyperplasia, atypical hyperplasia and carcinoma in situ. *Cancer* 1980; 45: 919–927.
15. Roa I, Araya J, Villaseca M et al. Preneoplastic lesions and gallbladder cancer: an estimate of the period required for progression. *Gastroenterology* 1996; 111: 232–236.
16. Roa I, Aretxabala XD, Roa J et al. Preneoplastic lesions in gallbladder cancer. *J Surg Oncol* 2006; 93: 615–623.
17. Yamagiwa H, Tomiyama H. Intestinal metaplasia–dysplasia–carcinoma sequence of the gallbladder. *Acta Pathol Jpn* 1986; 36: 989–997.
18. Duarte I, Llanos O, Domke H et al. Metaplasia and precursor lesions of gallbladder carcinoma. Frequency, distribution, and probability of detection in routine histologic samples. *Cancer* 1993; 72: 1878–1884.
19. Roa I, Araya J, Wistuba I et al. Epithelial lesions associated with gallbladder carcinoma. A methodical study of 32 cases. *Rev Med Chil* 1993; 121: 21–29.
20. Wistuba I, Gazdar A, Roa I et al. p53 protein overexpression in gallbladder carcinoma and its precursor lesions: an immunohistochemical study. *Hum Pathol* 1996; 27: 360–365.
21. Misra S, Chaturvedi A, Goel M et al. Overexpression of p53 protein in gallbladder carcinoma in North India. *Eur J Surg Oncol* 2000; 26: 164–167.
22. Kozuka S, Tsubone N, Yasui A et al. Relation of adenoma to carcinoma in the gallbladder. *Cancer* 1982; 50: 2226–2234.
23. Watanabe H, Date K, Itoi T et al. Histological and genetic changes in malignant transformation of gallbladder adenoma. *Ann Oncol* 1999; 10: 136–139.
24. Nakajo S, Yamamoto M, Tahara E. Morphometrical analysis of gallbladder adenoma and adenocarcinoma with reference to histogenesis and adenoma–carcinoma sequence. *Virchows Arch A Pathol Anat Histopathol* 1990; 417: 49–56.
25. Sasatomi E, Tokunaga O, Miyazaki K. Precancerous conditions of gallbladder carcinoma: overview of histopathologic characteristics and molecular genetic findings. *J Hepatobiliary Pancreat Surg* 2000; 7: 556–567.

26. Goldin R, Roa J. Gallbladder cancer: a morphological and molecular update. *Histopathology* 2009; 55: 218–229.
27. Pai RK, Mojtahed K. Mutations in the RAS/RAF/MAP kinase pathway commonly occur in gallbladder adenomas but are uncommon in gallbladder adenocarcinomas. *Appl Immunohistochem Mol Morphol* 2011; 19: 133–140.
28. Shukla P, Barreto S, Gupta P et al. Is there a role for estrogen and progesterone receptors in gall bladder cancer? *HPB (Oxford)* 2007; 9: 285–288.
29. Shukla P, Barreto S, Shrikhande S et al. Simultaneous gallbladder and bile duct cancers: revisiting the pathological possibilities. *HPB (Oxford)* 2008; 10: 48–53.
30. Chijiwa K, Tanaka M, Nakayama F. Adenocarcinoma of the gallbladder associated with anomalous pancreaticobiliary ductal junction. *Am Surg* 1993; 59: 430–434.
31. Wistuba II, Albores-Saavedra J. Genetic abnormalities involved in the pathogenesis of gallbladder carcinoma. *J Hepatobiliary Pancreat Surg* 1999; 6: 237–244.
32. Wistuba II, Gazdar AF. Gallbladder cancer: lessons from a rare tumour. *Nat Rev Cancer* 2004; 4: 695–706.
33. Castillo J, Garcia P, Roa J. Genetic alterations in preneoplastic and neoplastic injuries of the gallbladder. *Rev Med Chile* 2010; 138: 596–604.
34. Tazuma S, Kajiyama G. Carcinogenesis of malignant lesions of the gall bladder. The impact of chronic inflammation and gallstones. *Langenbecks Arch Surg* 2001; 386: 224–229.
35. Tanno S, Obara T, Fujii T et al. Epithelial hyperplasia of the gallbladder in children with anomalous pancreaticobiliary ductal union. *Hepatogastroenterology* 1999; 46: 3068–3073.
36. Hanada K, Itoh M, Fujii K et al. Pathology and cellular kinetics of gallbladder with an anomalous junction of the pancreaticobiliary duct. *Am J Gastroenterol* 1996; 91: 1007–1011.
37. Hanada K, Tsuchida A, Kajiyama G. Cellular kinetics and gene mutations in gallbladder mucosa with an anomalous junction of pancreaticobiliary duct. *J Hepatobiliary Pancreat Surg* 1999; 6: 223–228.
38. Roa EI, Munoz NS, Ibacache SG et al. Natural history of gallbladder cancer. Analysis of biopsy specimens. *Rev Med Chil* 2009; 137: 873–880.
39. Roa JC, Roa I, Correa P et al. Microsatellite instability in preneoplastic and neoplastic lesions of the gallbladder. *J Gastroenterol* 2005; 40: 79–86.
40. Lin HT, Liu GJ, Wu D et al. Metastasis of primary gallbladder carcinoma in lymph node and liver. *World J Gastroenterol* 2005; 11: 748–751.
41. Albores-Saavedra J, Chable-Montero F, Gonzalez-Romo MA et al. Adenomas of the gallbladder. Morphologic features, expression of gastric and intestinal mucins, and incidence of high-grade dysplasia/carcinoma in situ and invasive carcinoma. *Hum Pathol* 2012; 43: 1506–1513.
42. Rai R, Tewari M, Kumar M et al. p53: its alteration and gallbladder cancer. *Eur J Cancer Prev* 2011; 20: 77–85.
43. Yokoyama N, Hitomi J, Watanabe H et al. Mutations of p53 in gallbladder carcinomas in high-incidence areas of Japan and Chile. *Cancer Epidemiol Biomarkers Prev* 1998; 7: 297–301.
44. Moreno M, Pimentel F, Gazdar AF et al. TP53 abnormalities are frequent and early events in the sequential pathogenesis of gallbladder carcinoma. *Ann Hepatol* 2005; 4: 192–199.
45. Wistuba II, Sugio K, Hung J et al. Allele-specific mutations involved in the pathogenesis of endemic gallbladder carcinoma in Chile. *Cancer Res* 1995; 55: 2511–2515.
46. Feng Z, Chen J, Wei H et al. The risk factor of gallbladder cancer: hyperplasia of mucous epithelium caused by gallstones associates with p16/CyclinD1/CDK4 pathway. *Exp Mol Pathol* 2011; 91: 569–577.
47. Kim YT, Kim J, Jang YH et al. Genetic alterations in gallbladder adenoma, dysplasia and carcinoma. *Cancer Lett* 2001; 169: 59–68.
48. Saetta A, Lazaris AC, Davaris PS. Detection of ras oncogene point mutations and simultaneous proliferative fraction estimation in gallbladder cancer. *Pathol Res Pract* 1996; 192: 532–540.
49. Hanada K, Tsuchida A, Iwao T et al. Gene mutations of K-ras in gallbladder mucosae and gallbladder carcinoma with an anomalous junction of the pancreaticobiliary duct. *Am J Gastroenterol* 1999; 94: 1638–1642.
50. Matsubara T, Sakurai Y, Sasayama Y et al. K-ras point mutations in cancerous and noncancerous biliary epithelium in patients with pancreaticobiliary maljunction. *Cancer* 1996; 77: 1752–1757.
51. Itoi T, Watanabe H, Ajioka Y et al. APC, K-ras codon 12 mutations and p53 gene expression in carcinoma and adenoma of the gall-bladder suggest two genetic pathways in gall-bladder carcinogenesis. *Pathol Int* 1996; 46: 333–340.
52. Legan M, Luzar B, Marolt VF et al. Expression of cyclooxygenase-2 is associated with p53 accumulation in premalignant and malignant gallbladder lesions. *World J Gastroenterol* 2006; 12: 3425–3429.
53. Fumino S, Tokiwa K, Ono S et al. Cyclooxygenase-2 expression in the gallbladder of patients with anomalous arrangement of the pancreaticobiliary duct. *J Pediatr Surg* 2003; 38: 585–589.
54. Ghosh M, Kawamoto T, Koike N et al. Cyclooxygenase expression in the gallbladder. *Int J Mol Med* 2000; 6: 527–532.
55. Yoshida T, Sugai T, Habano W et al. Microsatellite instability in gallbladder carcinoma: two independent genetic pathways of gallbladder carcinogenesis. *J Gastroenterol* 2000; 35: 768–774.
56. Rai R, Sharma KL, Tiwari S et al. DCC (deleted in colorectal carcinoma) gene variants confer increased susceptibility to gallbladder cancer (Ref. No.: Gene-D-12-01446). *Gene* 2013; 518: 303–309.
57. Cha PC, Zembutsu H, Takahashi A et al. A genome-wide association study identifies SNP in DCC is associated with gallbladder cancer in the Japanese population. *J Hum Genet* 2012; 57: 235–237.
58. Wistuba II, Ashfaq R, Maitra A et al. Fragile histidine triad gene abnormalities in the pathogenesis of gallbladder carcinoma. *Am J Pathol* 2002; 160: 2073–2079.
59. Koda M, Yashima K, Kawaguchi K et al. Expression of Fhit, Mh1, and P53 protein in human gallbladder carcinoma. *Cancer Lett* 2003; 199: 131–138.
60. Knudson AG, Jr. Hereditary cancer, oncogenes, and antioncogenes. *Cancer Res* 1985; 45: 1437–1443.
61. Wistuba II, Maitra A, Carrasco R et al. High resolution chromosome 3p, 8p, 9q and 22q allelotyping analysis in the pathogenesis of gallbladder carcinoma. *Br J Cancer* 2002; 87: 432–440.
62. Chang HJ, Kim SW, Kim YT et al. Loss of heterozygosity in dysplasia and carcinoma of the gallbladder. *Mod Pathol* 1999; 12: 763–769.
63. Aarnio M, Sankila R, Pukkala E et al. Cancer risk in mutation carriers of DNA-mismatch-repair genes. *Int J Cancer* 1999; 81: 214–218.
64. Kohya N, Miyazaki K, Matsukura S et al. Deficient expression of O(6)-methylguanine-DNA methyltransferase combined with mismatch-repair proteins hMLH1 and hMSH2 is related to poor prognosis in human biliary tract carcinoma. *Ann Surg Oncol* 2002; 9: 371–379.
65. Mishra PK, Raghuram GV, Jatava SK et al. Frequency of genetic alterations observed in cell cycle regulatory proteins and microsatellite instability in gallbladder adenocarcinoma: a translational perspective. *Asian Pac J Cancer Prev* 2011; 12: 573–574.
66. Matsuda Y. Microsatellite instability meets gallbladder cancer. *J Gastroenterol* 2000; 35: 798–799.
67. Toledo C, Matus CE, Barraza X et al. Expression of HER2 and bradykinin B(1) receptors in precursor lesions of gallbladder carcinoma. *World J Gastroenterol* 2012; 18: 1208–1215.
68. Kamel D, Paakko P, Nuorva K et al. p53 and c-erbB-2 protein expression in adenocarcinomas and epithelial dysplasias of the gall bladder. *J Pathol* 1993; 170: 67–72.
69. Kim YW, Huh SH, Park YK et al. Expression of the c-erbB-2 and p53 protein in gallbladder carcinomas. *Oncol Rep* 2001; 8: 1127–1132.
70. Chaube A, Tewari M, Garbyal RS et al. Preliminary study of p53 and c-erbB-2 expression in gallbladder cancer in Indian patients manuscript id: 8962091628764582. *BMC Cancer* 2006; 6: 126.
71. Cabibi D, Licata A, Barresi E et al. Expression of cytokeratin 7 and 20 in pathological conditions of the bile tract. *Pathol Res Pract* 2003; 199: 65–70.
72. Chang HJ, Kim SW, Lee BL et al. Phenotypic alterations of mucins and cytokeratins during gallbladder carcinogenesis. *Pathol Int* 2004; 54: 576–584.
73. Chang YT, Hsu C, Jeng YM et al. Expression of the caudal-type homeodomain transcription factor CDX2 is related to clinical outcome in biliary tract carcinoma. *J Gastroenterol Hepatol* 2007; 22: 389–394.
74. Duval JV, Savas L, Banner BF. Expression of cytokeratins 7 and 20 in carcinomas of the extrahepatic biliary tract, pancreas, and gallbladder. *Arch Pathol Lab Med* 2000; 124: 1196–1200.

75. Harder J, Waiz O, Otto F et al. EGFR and HER2 expression in advanced biliary tract cancer. *World J Gastroenterol* 2009; 15: 4511–4517.
76. Kalekou H, Miliaras D. Cytokeratin 7 and 20 expression in gallbladder carcinoma. *Pol J Pathol* 2011; 62: 25–30.
77. Kaufman M, Mehrotra B, Limaye S et al. EGFR expression in gallbladder carcinoma in North America. *Int J Med Sci* 2008; 5: 285–291.
78. Lee CS, Pirdas A. Epidermal growth factor receptor immunoreactivity in gallbladder and extrahepatic biliary tract tumours. *Pathol Res Pract* 1995; 191: 1087–1091.
79. Li QL, Yang ZL, Liu JQ et al. Expression of CDX2 and hepatocyte antigen in benign and malignant lesions of gallbladder and its correlation with histopathologic type and clinical outcome. *Pathol Oncol Res* 2011; 17: 561–568.
80. Moon WS, Park HS, Lee H et al. Co-expression of cox-2, C-met and beta-catenin in cells forming invasive front of gallbladder cancer. *Cancer Res Treat* 2005; 37: 171–176.
81. Nakazawa K, Dobashi Y, Suzuki S et al. Amplification and overexpression of c-erbB-2, epidermal growth factor receptor, and c-met in biliary tract cancers. *J Pathol* 2005; 206: 356–365.
82. Ooi A, Suzuki S, Nakazawa K et al. Gene amplification of Myc and its coamplification with ERBB2 and EGFR in gallbladder adenocarcinoma. *Anticancer Res* 2009; 29: 19–26.
83. Sakamoto H, Mutoh H, Ido K et al. A close relationship between intestinal metaplasia and Cdx2 expression in human gallbladders with cholelithiasis. *Hum Pathol* 2007; 38: 66–71.
84. Sasaki M, Yamato T, Nakanuma Y et al. Expression of MUC2, MUC5AC and MUC6 apomucins in carcinoma, dysplasia and non-dysplastic epithelia of the gallbladder. *Pathol Int* 1999; 49: 38–44.
85. Shafizadeh N, Grenet JP, Sahai V et al. Epidermal growth factor receptor and HER-2/neu status by immunohistochemistry and fluorescence in situ hybridization in adenocarcinomas of the biliary tree and gallbladder. *Hum Pathol* 2010; 41: 485–492.
86. Wu XS, Akiyama Y, Igar T et al. Expression of homeodomain protein CDX2 in gallbladder carcinomas. *J Cancer Res Clin Oncol* 2005; 131: 271–278.
87. Xiong L, Yang Z, Yang L et al. Expressive levels of MUC1 and MUC5AC and their clinicopathologic significances in the benign and malignant lesions of gallbladder. *J Surg Oncol* 2012; 105: 97–103.
88. Yamato T, Sasaki M, Watanabe Y et al. Expression of MUC1 and MUC2 mucin core proteins and their messenger RNA in gallbladder carcinoma: an immunohistochemical and in situ hybridization study. *J Pathol* 1999; 188: 30–37.
89. Ghosh M, Kamma H, Kawamoto T et al. MUC1 core protein as a marker of gallbladder malignancy. *Eur J Surg Oncol* 2005; 31: 891–896.
90. Kim HJ, Kim SH, Chae GB et al. Increased expression of mucin 5AC mRNA and decreased expression of epidermal growth-factor receptor mRNA in gallstone patients. *Tohoku J Exp Med* 2008; 214: 139–144.
91. Scaltriti M, Baselga J. The epidermal growth factor receptor pathway: a model for targeted therapy. *Clin Cancer Res* 2006; 12: 5268–5272.
92. Kim YS, Gum J, Jr, Brockhausen I. Mucin glycoproteins in neoplasia. *Glycoconj J* 1996; 13: 693–707.
93. Reis CA, David L, Correa P et al. Intestinal metaplasia of human stomach displays distinct patterns of mucin (MUC1, MUC2, MUC5AC, and MUC6) expression. *Cancer Res* 1999; 59: 1003–1007.
94. Chuang SC, Hsi E, Lee KT. Mucin genes in gallstone disease. *Clin Chim Acta* 2012; 413: 1466–1471.
95. Finzi L, Barbu V, Burgel PR et al. MUC5AC, a gel-forming mucin accumulating in gallstone disease, is overproduced via an epidermal growth factor receptor pathway in the human gallbladder. *Am J Pathol* 2006; 169: 2031–2041.
96. Vilkin A, Nudelman I, Morgenstern S et al. Gallbladder inflammation is associated with increase in mucin expression and pigmented stone formation. *Dig Dis Sci* 2007; 52: 1613–1620.
97. Boonla C, Wongkham S, Sheehan J et al. Prognostic value of serum MUC5AC mucin in patients with cholangiocarcinoma. *Cancer* 2003; 98: 1438–1443.
98. Park S, Roh S, Kim Y et al. Expression of MUC1, MUC2, MUC5AC and MUC6 in cholangiocarcinoma: prognostic impact. *Oncol Rep* 2009; 22: 649–657.
99. Natarajan S, Xu F, Gilchrist K et al. Cytokeratin is a superior marker for detection of micrometastatic biliary tract carcinoma. *J Surg Res* 2005; 125: 9–15.
100. Yokoyama N, Shirai Y, Hatakeyama K. Immunohistochemical detection of lymph node micrometastases from gallbladder carcinoma using monoclonal anticytokeratin antibody. *Cancer* 1999; 85: 1465–1469.
101. Tajima Y, Tomioka T, Ikematsu Y et al. Immunohistochemical demonstration of cytokeratin is useful for detecting micrometastatic foci from gallbladder carcinoma in regional lymph nodes. *Jpn J Clin Oncol* 1999; 29: 425–428.
102. Kim HS, Kim GY, Lim SJ et al. Reduced expression of Raf-1 kinase inhibitory protein is a significant prognostic marker in patients with gallbladder carcinoma. *Hum Pathol* 2010; 41: 1609–1616.
103. Li J, Wu T, Lu J et al. Immunohistochemical evidence of the prognostic value of hedgehog pathway components in primary gallbladder carcinoma. *Surg Today* 2012; 42: 770–775.
104. Li Q, Yang Z. Expression of phospho-ERK1/2 and PI3-K in benign and malignant gallbladder lesions and its clinical and pathological correlations. *J Exp Clin Cancer Res* 2009; 28: 65.
105. Liu DC, Yang ZL. Overexpression of EZH2 and loss of expression of PTEN is associated with invasion, metastasis, and poor progression of gallbladder adenocarcinoma. *Pathol Res Pract* 2011; 207: 472–478.
106. Liu DC, Yang ZL, Jiang S. Identification of PEG10 and TSG101 as carcinogenesis, progression, and poor-prognosis related biomarkers for gallbladder adenocarcinoma. *Pathol Oncol Res* 2011; 17: 859–866.
107. Wang W, Yang ZL, Liu JQ et al. Identification of CD146 expression, angiogenesis, and lymphangiogenesis as progression, metastasis, and poor-prognosis related markers for gallbladder adenocarcinoma. *Tumour Biol* 2012; 33: 173–182.
108. Yang L, Lan S, Liu J et al. Expression of MK-1 and RegIV and its clinicopathological significances in the benign and malignant lesions of gallbladder. *Diagn Pathol* 2011; 6: 100.
109. Yang Z, Xiong L, Huang S et al. Expression of VHL and HIF-1alpha and their clinicopathologic significance in benign and malignant lesions of the gallbladder. *Appl Immunohistochem Mol Morphol* 2011; 19: 534–539.
110. Zou Q, Xiong L, Yang Z et al. Expression levels of HMGA2 and CD9 and its clinicopathological significances in the benign and malignant lesions of the gallbladder. *World J Surg Oncol* 2012; 10: 92.
111. Roa I, de Arexabala X, Lantadilla S et al. ERCC1 (excision repair cross-complementing 1) expression in pT2 gallbladder cancer is a prognostic factor. *Histol Histopathol* 2011; 26: 37–43.
112. Hobbs SS, Goettel JA, Liang D et al. TNF transactivation of EGFR stimulates cytoprotective COX-2 expression in gastrointestinal epithelial cells. *Am J Physiol Gastrointest Liver Physiol* 2011; 301: G220–G229.
113. Gray T, Nettesheim P, Loftin C et al. Interleukin-1beta-induced mucin production in human airway epithelium is mediated by cyclooxygenase-2, prostaglandin E2 receptors, and cyclic AMP-protein kinase A signaling. *Mol Pharmacol* 2004; 66: 337–346.
114. Nadel JA. Role of epidermal growth factor receptor activation in regulating mucin synthesis. *Respir Res* 2001; 2: 85–89.
115. Vishnoi M, Pandey SN, Choudhuri G et al. IL-1 gene polymorphisms and genetic susceptibility of gallbladder cancer in a north Indian population. *Cancer Genet Cytogenet* 2008; 186: 63–68.
116. Sinicrope FA, Lemoine M, Xi L et al. Reduced expression of cyclooxygenase 2 proteins in hereditary nonpolyposis colorectal cancers relative to sporadic cancers. *Gastroenterology* 1999; 117: 350–358.
117. Karnes WE, Jr., Shattuck-Brandt R, Burgart LJ et al. Reduced COX-2 protein in colorectal cancer with defective mismatch repair. *Cancer Res* 1998; 58: 5473–5477.
118. Barreto SG, Haga H, Shukla PJ. Hormones and gallbladder cancer in women. *Indian J Gastroenterol* 2009; 28: 126–130.
119. Gerlinger M, Rowan AJ, Horswell S et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med* 2012; 366: 883–892.
120. Yap TA, Gerlinger M, Futreal PA et al. Intratumor heterogeneity: seeing the wood for the trees. *Sci Transl Med* 2012; 4: 127ps110.

121. Watson P, Vasen HF, Mecklin JP et al. The risk of extra-colonic, extra-endometrial cancer in the Lynch syndrome. *Int J Cancer* 2008; 123: 444–449.
122. Kim ET, Namgung H, Shin HD et al. Oncologic manifestations of neurofibromatosis type 1 in Korea. *J Korean Surg Soc* 2012; 82: 205–210.
123. Lowenfels AB, Maisonneuve P, Boyle P et al. Epidemiology of gallbladder cancer. *Hepatogastroenterology* 1999; 46: 1529–1532.
124. Hemminki K, Li X. Familial liver and gall bladder cancer: a nationwide epidemiological study from Sweden. *Gut* 2003; 52: 592–596.
125. Subbiah IM, Subbiah V, Tsimberidou AM et al. Targeted therapy of advanced gallbladder cancer and cholangiocarcinoma with aggressive biology: eliciting early response signals from phase 1 trials. *Oncotarget* 2013; 4: 156–165.
126. Subbiah IM, Subbiah V, Tsimberidou AM et al. Targeted therapy of advanced gallbladder cancer and cholangiocarcinoma with aggressive biology: eliciting early response signals from phase 1 trials. *Oncotarget* 2013; 4: 153–162.
127. Caldw Pilgrim CH, Groeschl RT, Quebbeman EJ et al. Recent advances in systemic therapies and radiotherapy for gallbladder cancer. *Surg Oncol* 2013; 22: 61–67.